Mercury and Methyl Mercury in California Fish, Water and Sediment: the Importance of Ecosystem Factors

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1	Summary
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3	Past anthropogenic activities in the state of California have led to an increase in levels of
4	mercury found in many of the state's freshwater fish species. The Surface Water Ambient
5	Monitoring Program- Bioaccumulation Oversight Group (SWAMP-BOG), funded by the
6	California State Water Resources Control Board, conducted a study examining fish-tissue
7	mercury concentrations in various lakes and reservoirs throughout the state. However no samples
8	were analyzed for mercury or methyl mercury at any of the locations sampled. This study,
9	funded by the State Water Resources Control Board for the development of methyl mercury
10	bioaccumulation factors, served to supplement the SWAMP-BOG data by collecting water data
11	to address the factors controlling mercury accumulation in fish. This included the calculation of
12	methyl mercury bioaccumulation factors, collection of ancillary water measurements to help
13	explain methyl mercury concentrations in water and fish, and developing correlations between
14	the fish and methyl mercury water data.
15	Mercury lake concentrations varied considerably among lakes especially during stratified
16	regimes where noticeable increases occurred in both near surface and bottom water methyl
17	mercury concentrations. There is some evidence to suggest buildup of aqueous methyl mercury
18	may be occurring in the thermocline in some lakes and which could be associated with lake
19	oxygen levels. These lakes may benefit by the installation of pumps or bubblers to reduce methyl
20	mercury concentrations in fish. There is also some evidence that increasing phytoplankton
21	concentrations in lakes may reduce methyl mercury concentrations in fish as well. There were
22	several factors that stand out as potential controlling factors that influence mercury
23	concentrations in fish. These include methyl mercury in near surface waters and total mercury in
24	sediments. To determine potential sources of methyl mercury in near surface waters however
25	will require mass balance studies in each lake.
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32 Introduction

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California's historic gold and mercury (Hg) mining has led to a significant amount of 34 mercury being released into the environment (Wiener and Suchanek 2008). As a result, many of 35 California's lakes and rivers have become impaired due to high levels of mercury found in local 36 populations of fish (Davis et al. 2009; Melwani et al. 2009). While most of the mercury in 37 California exists as inorganic mercury (Hg^{II} and Hg⁰), the organic and highly toxic form is 38 methyl mercury (MeHg). Bacteria convert mercury to methyl mercury which is concentrated by 39 microorganisms and subsequently biomagnified in food webs (Wiener et al. 2003). Nearly all of 40 the mercury accumulated by fish and higher trophic levels is methyl mercury (Bloom 1992) 41 which, when consumed, can adversely affect human health and wildlife. 42 Human consumption of fish species containing methyl mercury, even in low 43 concentrations, can adversely affect the nervous, renal, immune, and reproductive systems in the 44

concentrations, can adversely affect the hervous, renal, immune, and reproductive systems in th
adult, child, and developing fetus stages of human life (Zahir et al. 2005). Methyl mercury has
also been shown to impair foraging efficiency, adversely affect endocrine systems, and
reproduction in fish (Drevnick and Sandheinrich 2003; Hammerschmidt et al. 2002) and birds
(Brasso and Cristol 2008; Schwarzbach et al. 2009). Considering the impact mercury can have
on both humans and wildlife, addressing contaminant levels of mercury has been the focus of
several studies in California.

Recently, the state of California's Surface Water Ambient Monitoring Program Bioaccumulation Oversight Group (SWAMP-BOG) conducted a study examining fish-tissue mercury concentrations in various lakes and reservoirs. Out of 152 lakes sampled, 74% were above the state's advisory tissue level (ATL) of 3 servings per week for mercury and 26% were above the no consumption range (>440 ppb; (Davis et al. 2009). However, it was unclear as to why some of these lakes had elevated levels of tissue mercury when past anthropogenic activities, such as mining did, not appear to be a factor.

Therefore, the objectives of our study were to (1) collect water methyl mercury data to adequately characterize the concentrations in a sub-set of lakes used in the SWAMP-BOG study, (2) collect ancillary parameter water data to help explain methyl mercury concentrations in water and fish, (3) develop correlations between the fish and mercury, and (4) calculate methyl mercury bioaccumulation factors (BAFs). Additional funding for the study was provided by the

63	Regional Water Quality Control Board (RWQCB) in Sacramento to supplement their ongoing
64	Total Maximum Daily Load (TMDL) efforts. This study will also provide supplemental data to
65	the Office of Environmental Health Hazard Assessment (OEHHA) for regulatory purposes to
66	assist in developing fish consumption advisories . The use of the data allows OEHHA and
67	RWQCB personnel to better address factors affecting methylation, suggest best management
68	practices (BMPs), and implement control measures as necessary.
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70	Methods
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72	Lake Selection
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74	Lakes were chosen based on their relative mercury concentrations in fish-tissue,
75	primarily largemouth bass (data provided by SFEI, taken from the SWAMP-BOG study), and
76	accessibility. Additional consideration was made to include lakes that were part of an ongoing
77	RWQCB-TMDL effort. The final study design included 28 lakes (Table 1). Twenty-two of these
78	lakes were sampled bi-monthly or monthly depending on RWQCB-TMDL programs from
79	August 2008 through October 2009 and some of these lakes had, within in them, multiple
80	stations were water and sediment was collected (See Table 1). The additional six lakes (Bass,
81	Britton, Butt Valley, Rollins, New Melones, and Paradise) were sampled just once in September
82	2009, as part of RWQCB 5's TMDL effort. These lakes had associated fish mercury data as well.
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84	Sample Collection
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86	All water and sediment sampling was conducted by RWQCB and California Department
87	of Fish and Game-Marine Pollution Studies Laboratory (CDFG-MPSL). A multi-parameter
88	(probe)YSI 600 xL sonde measuring temperature, conductivity, pH, and oxygen was used to
89	determine lake stratification prior to any water collections. Probe measurements were taken in 3-
90	meter increments from the surface to near-bottom (50 ft. max in depth; RWQCB 3, 5, & 8) at all
91	lakes and recorded. During the summer period, when most of the lakes were determined to be
92	consistently stratified (Jun-Sept), water was collected in epilimnion (near surface) and

93 hypolimnion (below thermocline; bottom) regions of each lake using a pre-cleaned (5% HCl)

94 Wildco Teflon Kemmerer sampler. During the winter period (Oct-Apr), when most of the lakes were determined to be well mixed, only a near surface sample was collected, with the exception 95 96 of RWQCB 5 which sampled both near surface and bottom water throughout the study. All samples collected for aqueous mercury (total and methyl) were collected unfiltered and using a 97 clean hands dirty hands techniques (Mpsl-Dfg_Fieldsop_V1.0 2007). The purpose of this study 98 design was to characterize unfiltered aqueous mercury concentrations on a temporal scale and to 99 examine any effects stratification may have on the distribution of unfiltered mercury in the water 100 column. 101

Near surface water samples were analyzed for methyl mercury, Chlorophyll *a*, sulfates (SO4), and dissolved organic carbon (DOC) and deep water samples were only analyzed for methyl mercury. An additional water sample was collected for total mercury analysis on two separate sampling events (stratified and non-stratified lake type) coinciding with the regular sampling of lakes. One sediment sample was collected in the deepest depositional area of each lake using a Van Veen grab sampler (0.5 m^2). All sediment samples were analyzed for total mercury and total organic carbon (TOC).

Big Bear Lake, Lake Englebright, Thermalito Forebay, and Lake Hemet had no
largemouth bass mercury data so an additional effort was put forth to collect fish at these lakes.
CDFG-MPSL staff, using a Smith-Root Electrofishing boat, put forth an effort to collect fish at
each of these lakes. As a result, largemouth bass were collected at Lake Hemet and Big Bear
Lake while spotted bass were collected at Lake Englebright. However, no fish were collected
from Thermalito Forebay.

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116 Sample Custody

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All samples were placed on ice or frozen and shipped overnight back to Moss Landing Marine Laboratories for analysis unless CDFG-MPSL personnel were able to drive them back to the laboratory. Standard two day hold times before acidification applied to all mercury water samples.

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125 Analysis

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The analytical and collecting techniques are identical to those used in current SWAMP
programs. The investigators are involved in several SWAMP funded projects and quality
assurance/quality control provisions of the study were identical to those in the SWAMP Quality
Assurance Project Plan (QAPP). All protocols are available upon request.

All methyl mercury water samples were analyzed according to EPA 1630. They were distilled to separate methyl mercury from the water matrix (Horvat 1993). An ethylating agent was added to each sample to form a volatile methyl-ethyl mercury derivative, and then purged onto graphite carbon traps as a means of preconcentration and interference removal. The sample was then isothermally chromatographed, pyrolitically broken down to elemental mercury, and detected using a cold vapor fluorescence detector. Sample results are corrected for distillation efficiency.

Total mercury samples were analyzed using Modified EPA 1631, Revision E: Mercury in
Water by Oxidation, Purge and Trap, and Cold Vapor Atomic Fluorescence Spectrometry (Usepa
2002). Sediment samples were prepared by cold aqua-regia digestion (MPSL-107) and analyzed
using a Flow Injection Mercury System (FIMS; MPSL-103). TOC in sediments were measured
as percent loss on ignition (LOI).

Tissue-mercury samples were analyzed by USEPA method 7473 for the SWAMP-BOG program. Samples were dissected using standard clean procedures and analyzed on a Milestone Direct Mercury Analyzer (DMA-80). Briefly, the method involves a drying step followed by combustion, purging, trapping on gold, desorbtion, and AA detection. All fish samples used in this study were analyzed as individuals.

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149 Statistical Analysis

Data were transformed to meet assumptions of statistical tests and all tests were performed using SPSS or SYSTAT software packages. Values that fell below detection limits were set to one half the reporting limit (R.L.). Anything less than 0.05 was considered to be statistically significant.

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156	Bioaccumulation Factors
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158	Methyl mercury BAFs were calculated for each lake using equation (1).
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160	$BAF_T^t = \frac{C_t}{C_w}$
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162	$C_t = Total$ concentration of chemical in fish tissue.
163	$C_w = Total$ concentration of chemical in water.
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165	All BAFs reported in this document were calculated using the above equation and are
166	expressed in units of L•Kg ⁻¹ . BAF values were calculated using either the geometric mean or
167	average of unfiltered aqueous methyl mercury concentrations (Sanborn 2006; USEPA 2003).
168	Total mercury tissue concentrations were calculated using a length-tissue mercury regression and
169	concentrations were normalized to 350 mm.
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171	Results
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173	Mercury in study lakes
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175	Average near surface methyl mercury concentrations varied from $< RL$ (Expressed as $\frac{1}{2}$
176	the RL, 0.010 ng•L ⁻¹) to 0.215 ng•L ⁻¹ and bottom methyl mercury varied from $<$ RL to 0.351
177	ng•L ⁻¹) among the stations in the study. Total mercury in sediments varied from $<$ RL to 3.09
178	$\mu g^{\bullet}g^{-1}$, the highest being found at the Lake Nacimiento Las Tablas station. This was not
179	surprising considering an abandoned mercury mine (Klau/Buena Vista), a designated EPA
180	superfund site, discharges directly into Las Tablas Creek, which flows into Nacimiento.
181	Table 2 summarizes the average, over the study period, of samples collected at each station.
182	Average methyl mercury concentrations within each lake and normalized (350 mm)
183	mercury tissue concentrations were used for the purpose of calculating BAFs (Table 3).
184	Calculations were made using data from the entire study period. Average mercury concentration

in bass species varied considerably among lakes $(91 - 1314 \text{ ng} \cdot \text{g}^{-1})$ as well as average methyl mercury concentrations (<RL - 0.158 ng \cdot \text{g}^{-1}).

187 Observed methyl mercury concentrations among study lakes varied more in stratified (summer) versus non-stratified (winter) regimes (Figure 1a). A large portion of the variability 188 had to do with the distribution of aqueous methyl mercury concentrations in the water column 189 during lake stratification (Figure 1b). Most notably, Lake San Antonio and Lake Hemet average 190 bottom concentrations were around two orders of magnitude higher than their average near 191 surface water concentrations during summer months (0.708 and 0.368 versus 0.051 and 0.020 192 ng•L, respectively). However, largemouth bass tissue concentrations were among the lowest in 193 the study for these two lakes (302 and 177 ppb) and below the no consumption limit of 440 ppb. 194 Lake Pillsbury and Lake Nacimiento had the highest average near surface aqueous methyl 195 mercury concentrations during summer months (0.163 and 0.089 ng•L) and the highest 196 largemouth bass tissue concentrations in the study (1,314 and 1,236 ppb). The discrepancy 197 between near surface and bottom water in Pillsbury and Nacimiento was not as large as in Lake 198 San Antonio and Hemet. Lake Pillsbury's average near surface methyl mercury concentrations 199 200 were only about one and a half times higher than the average bottom water concentrations (0.163) and 0.121 ng•L) and roughly a 2-fold difference between near surface and bottom water in Lake 201 Nacimiento (0.089 and 0.043 ng•L). 202

Additional water samples were collected from Lake Nacimiento and Lake San Antonio 203 204 in September 2009 to further profile the distribution of methyl mercury in the water column (Figure 2). The methyl mercury profile taken at Lake San Antonio was similar to the trend seen 205 in the monthly summer collections: near surface concentrations were relatively low while bottom 206 water concentrations were high. Lake Nacimiento however, had a rather large methyl mercury 207 spike $(3.73 \text{ ng} \cdot \text{L}^{-1})$ near the oxygen minimum. We examined six additional profiles (see methods 208 for the list of lakes) to ascertain whether this mid-water methyl mercury phenomena occurred in 209 210 other lakes as well. Unfortunately, these lakes had either de-stratified by the time samples were collected or the methylmercury concentrations were too low (within 3x R.L.) to adequately 211 212 assess whether the same phenomena had occurred.

Total mercury in water was measured twice (1 summer /1 winter) at each study lake with the exception of Pillsbury, Nacimiento, and San Antonio where only one summer collection was made. Aqueous total mercury concentrations were similar to the pattern observed in the aqueous 216 methyl mercury data (Figure 1). Lake Nacimiento and Camp Far West were outliers among

study lakes with respect to total mercury concentrations. Furthermore, Camp Far West had the

third highest largemouth bass tissue concentrations in the study (843 ppb) behind Pillsbury andNacimiento.

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221 Ancillary measurements and correlations with fish tissue mercury

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Variables measured in the study were examined using a Pearson correlation to ascertain
which factors may potentially be important in driving bioaccumulation in largemouth bass
(LMB) only. Averaged summer datasets were compared against averaged winter datasets to
determine if there were any temporal trends among the mercury variables relative to LMB
mercury concentrations (n = 17; Table 4).

Overall, total mercury in sediment had the highest correlation to LMB tissue mercury concentrations (r = 0.709; p < 0.01) and explained roughly 50% of the variability in LMB mercury concentrations ($R^2 = 0.503$; p < 0.01; Figure 3). Averaged summer near surface methyl mercury concentrations were moderately correlated to LMB (r = 0.473; p = 0.055) while winter concentrations were weakly correlated at best(r = 0.235; p = 0.363). Total mercury concentrations were moderately correlated to LMB in the winter (r = 0.433; p = 0.094) and summer near surface/bottom concentrations weakly correlated (r < 0.400; p > 0.10).

Summer DOC, SO4, chl-*a*, and specific conductivity ancillary measurements had a significant and moderate correlation (negative) with LMB mercury concentrations (r > -0.500; p < 0.05). The same results held true in the winter, with the exception of DOC, which had no significant correlation with largemouth bass during this period. All of these variables, regardless of summer or winter periods, typically had a strong correlation among each other (r > 0.700; p 0.01).

Among the mercury variables, total mercury in sediments was the only variable that had a significantly negative correlation with both chl-*a* and SO4 (r = -0.547, -0.493 respectively; p <0.05) in the summer period and only with chl-*a* during the winter period. Near surface methyl mercury concentrations were moderately correlated with DOC (r = 0.533; p = 0.028) and pH (r =0.563; p = 0.563) in the winter period and had no significant correlations with any of the other ancillary measurements.

247 We also examined the relationship between mercury in lakes and all bass species collected with respect to general areas of the state (Table 5). In general, Coast Range lakes 248 249 (RWQBs 1 and 3) were high in LMB, methyl, and total mercury in surface waters. Additionally these lakes were high in sediment total mercury and had a high degree of stratification. The 250 Sierra Nevada Lakes (RWQB 5) were medium to high in LMB mercury (for the most part) and 251 low to medium in the degree of stratification. The southern California lakes (RWQCB 8) were 252 253 low in LMB mercury, low to medium in methyl and total mercury surface waters and variable in stratification. In general, mercury concentrations were highest in Coast Range lakes, medium in 254 the Sierra's, and lowest in southern California. However, there were exceptions in each, which 255 illustrates the complexity of the lakes and supports the concept that there are multiple processes 256 occurring at each lake that control the concentration of mercury in bass species. 257

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- 259 Discussion
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261 Oxygen and stratification

Although there was no significant correlation between the degree of stratification and LMB mercury among all study lakes, there was however a significant correlation between the degree of stratification and LMB mercury concentration in the Sierra Nevada Lakes (r^2 = 0.68, p < 0.01; Figure 4). We are currently examining possible reasons attributed to why this only seen in the Sierra Nevada lakes.

Two of our study lakes, San Antonio and Hemet, had significant levels of hypolimnetic 267 methyl mercury associated with low dissolved oxygen and six of the lakes had > 65% change in 268 oxygen concentrations between surface and bottom waters but little evidence of a buildup of 269 270 hypolimnetic methyl mercury. Studies have shown methyl mercury formation in anoxic bottom water can be attributed to in situ processes and may not necessarily be controlled by source 271 inputs (Eckley et al. 2005). Methyl mercury in fish has also been attributed to methyl mercury in 272 bottom water in other areas such as Davis Creek Reservoir (Slotton et al. 1995) and lakes in the 273 274 Guadalupe Watershed near San Jose California. Both San Antonio and Hemet had elevated surface methyl mercury concentrations in mid to late summer months when stratification was the 275 greatest. This suggests that in situ processes may be controlling bottom water methyl mercury 276

277 concentrations in these lakes. However, at neither of these lakes were mercury tissue

- 278 concentrations elevated.
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280 Near surface water methyl and total mercury concentrations

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There was a moderate correlation between methyl mercury in surface waters and LMB in 282 the summer (r = 0.47) and a lesser correlation between total mercury in surface waters and LMB 283 in the summer and winter (r = 0.36 - 0.43 respectively). In the two study lakes that did have the 284 highest tissue mercury concentrations, Pillsbury and Nacimiento, near surface methyl mercury 285 concentrations were also the highest among lakes indicating in these two lakes epilimnetic 286 methyl mercury may be causing high tissue levels. Others have found methyl mercury and total 287 mercury in surface waters to be an important factor for influencing the bioaccumulation of 288 mercury in fish (Wiener et al. 2006). 289

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291 Total mercury in sediments

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Total mercury in sediments was correlated to LMB mercury (r = 0.71) and total mercury 293 in surface waters in summer and winter periods (r = 0.48-0.51). Total mercury in sediments 294 was used as a proxy for the presence of mercury in the watershed due to mining activities or 295 296 natural deposits of cinnabar. The total mercury in sediments was also correlated to methyl mercury in surface waters indicating the methyl mercury may be formed in the sediments and 297 fluxed up into the surface waters or alternatively methyl and total mercury could be co-occurring 298 in water brought into the lakes via tributaries. To determine whether methyl mercury in water is 299 300 coming from in place sediments or the tributaries is not possible with this data set and would need follow-up mass balance studies to make this assessment. In one study at Folsom Reservoir, 301 302 the Sacramento Regional Water Quality Control Board determined the methyl mercury concentrations could not be accounted for by tributary inputs alone and there must be some 303 304 within lake production (Chris Foe, personal communication). 305

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308 Conclusion

310	The concentrations of mercury in LMB are correlated (for the most part) to methyl
311	mercury and total mercury in surface waters, and total mercury in sediments. In the Sierra
312	Nevada Range the concentration of mercury in LMB was also correlated to degree of lake
313	stratification. There were four lakes that exhibited a high degree (>65%) of stratification and
314	high levels of mercury in LMB and would be candidates for destratification using pumps or
315	bubblers. These include Lakes Pillsbury, Mendicino, Nacimiento, and Camp Far West.
316	Destratification best management practices have been used successfully in lakes in the
317	Guadalupe Watershed near San Jose California to lower the concentrations of mercury in fish.
318	The concentrations of mercury in LMB were also negatively correlated with chlorophyll
319	a in surface waters indicating stimulating growth of phytoplankton may reduce mercury in fish.
320	Chlorophyll a has been shown in other studies to be negatively correlated to LMB and Clams in
321	the Delta (Foe, personal communication). Others have found algal blooms reduce the uptake of
322	methyl mercury in freshwater food webs (Pickhardt et al., 2002).
323	Both methyl mercury in surface waters and total mercury in sediment are correlated to mercury
324	in LMB. To determine whether control measures to reduce methyl mercury loadings to the lake
325	by reducing inputs from the tributaries would be successful mass balance studies would be
326	necessary to determine the relative amounts of methyl mercury coming from the tributaries and
327	within lake sources.
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369	References
370	
371	Bloom, N. S. 1992. On the chemical form of mercury in edible fish and marine vertebrate tissue.
372	Can. J. Aquat. Sci. 49.
373	Brasso, R., and D. Cristol. 2008. Effects of mercury exposure on the reproductive success of tree
374	swallows (Tachycineta bicolor). Ecotoxicology 17: 133-141.
375	Davis, J. A. and others 2009. Contaminants in Fish From California Lakes and Reservoirs:
376	Technical Report on Year One of a Two-Year Screening Study. A Report of the Surface
377	Water Ambient Monitoring Program (SWAMP). California State Water Resources
378	Control Board, Sacramento, CA.
379	Drevnick, P. E., and M. B. Sandheinrich. 2003. Effects of Dietary Methylmercury on
380	Reproductive Endocrinology of Fathead Minnows. Environmental Science & Technology
381	37: 4390-4396.
382	Eckley, C. S., C. J. Watras, H. Hintelmann, K. Morrison, A. D. Kent, and O. Regnell. 2005.
383	Mercury methylation in the hypolimnetic waters of lakes with and without connection to
384	wetlands in northern Wisconsin. Can. J. Fish. Aquat. Sci. 62(2): 400-411.
385	Hammerschmidt, C. R., M. B. Sandheinrich, J. G. Wiener, and R. G. Rada. 2002. Effects of
386	Dietary Methylmercury on Reproduction of Fathead Minnows. Environmental Science &
387	Technology 36: 877-883.
388	Horvat, M., Liang, L., & Bloom, N. S 1993. Comparison of distillation with other current
389	isolation methods for the determination of methyl mercury compounds in low level
390	environmental samples. part II. water. Analytica Chimica Acta 282(1): 153-168.

- Melwani, A. R. and others 2009. Spatial trends and impairment assessment of mercury in sport
 fish in the Sacramento-San Joaquin Delta watershed. Environmental Pollution 157: 31373149.
- ---. 2010. Factors Influencing the Bioaccumulation of Methylmercury in Largemouth Bass from
 California Lakes and Reservoirs, Draft Report. .
- 396 Mpsl-Dfg_Fieldsop_V1.0. 2007. Marine Pollution Studies Laboratory Department of Fish and
- 397 Game (MPSL-DFG) Standard Operating Procedures (SOPs) for Conducting Field
- 398 Measurements and Field Collections of Water and Bed Sediment Samples in the Surface
- 399 Water Ambient Monitoring Program (SWAMP).
- 400 Pickhardt, P.C., C. Folt, C. Chen, B. Klaue, and J.D. Blum. 2002. Algal blooms reduce the
- 401 uptake of toxic methylmercury in freshwater food webs. PNAS: 99(7): 4479-4423.
- Sanborn, J. R., Brodberg, R.K. 2006. Evaluation of Bioaccumulation Factors and Translators
 for Methylmercury. *In* O. o. E. H. H. Assessment [ed.].
- 404 Schwarzbach, S. E., J. D. Albertson, C. M. Thomas, and D. B. Lank. 2009. Effects of predation,
- flooding, and contamination on reproductive success of California Clapper Rails (Rallus
 longirostris obsoletus) in San Francisco Bay. The Auk 123: 45-60.
- 407 Simoneau, M., M. Lucotte, S. Garceau, D. Laliberte. 2005. Fish Growth Rates Modulate

408 Mercury Concentrations in walleye (Sander vitreus) from eastern Canadian Lakes.
409 Environ. Res. 98: 73-82.

- 410 Slotton, D.G., J.E. Reuter, and C.R. Goldman. 1995. Mercury Uptake Patterns of Biota in a
- 411 Seasonally Anoxic Northern California Reservoir. Water, Air and Soil Pollution 80: 841412 850.
- 413

- Fluorescence Spectrometry, Revision E. . US Environmental Protection Agency. 415
- ---. 2003. Methodology for Deriving Ambient Water Quality Criteria for the Protection of 416
- 417 Human Health (2000). Technical Support Document Volume 2: Development of National
- Bioaccumulation Factors. In O. o. Water [ed.]. 418
- Wiener, J. G. and others 2006. Mercury in Soils, Lakes, and Fish in Voyageurs National Park 419
- (Minnesota): Importance of Atmospheric Deposition and Ecosystem Factors. 420
- Environmental Science & Technology 40: 6261-6268. 421
- Wiener, J. G., D. P. Krabbenhoft, G. H. Heinz, and A. M. Scheuhammer. 2003. Ecotoxicology of 422 mercury, Second ed. Lewis Publishers. 423
- Wiener, J. G., and T. H. Suchanek. 2008. The basis for ecotoxicological concern in aquatic 424 ecosystems contaminated by historic mercury mining. Ecological Applications 18: A3-425
- A11. 426
- Zahir, F., S. J. Rizwi, S. K. Haq, and R. H. Khan. 2005. Low dose mercury toxicity and human 427 health. Environmental Toxicology and Pharmacology 20: 351-360.

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RWQCB	Lake	Station	Latitude	Longitude	
1	Sonoma	Smith Creek	38.7398	-123.0722	
1	Sonoma	Dam	38.7205	-123.0205	
1	Mendocino	Dam	39.2009	-123.1754	
3	Mendocino	Russian River	39.2327	-123.1739	
3	Pillsbury	Dam	39.4229	-122.9547	
5	Pillsbury	Eel River	39.4126	-122.9242	
5	Naciemento	Dam	35.7629	-120.9030	
5	Naciemento	Las Tablas	35.7088	-120.9514	
5	San Antonio	Marina	35.8702	-121.0101	
5	San Antonio	Delta	35.8912	-121.0598	
5	Lake Engelbright		39.2519	-121.2705	
5	Thermalito Afterbay		39.4897	-121.6801	
5	Thermalito Forebay		39.5169	-121.6204	
5	Lake Oroville		39.5482	-121.4764	
5	Folsom Lake		38.7199	-121.1466	
5	Lake Natomas		38.6370	-121.2178	
5	Don Pedro		37.7144	-120.3942	
5	Lake McClure		37.5924	-120.2632	
5	Lake McSwain		37.5187	-120.2992	
5	San Luis Resrvoir		37.0609	-121.1024	
5	Oneil Forebay		37.0814	-121.0526	
5	Camp Far West		39.0339	-121.2831	
8	Big Bear		34.2476	-116.9567	
8	Irvine	Dam	33.7824	-117.7247	
8	Irvine	Santiago Flats	33.7755	-117.7045	
8	Perris	Dam	33.8556	-117.1815	
8	Perris	Allesandro	33.8648	-117.1714	
8	Hemet		33.6648	-116.7036	
8	Elsinore		33.6562	-117.3518	
Additional L	akes sampled one time	in September 2009			
5	Bass Lake*	37.3139	-119.5474		
5	Lake Britton*		41.0276	-121.6426	
5	Butt Valley Reservoir	*	40.1459	-121.1739	
5	Rollins Reservoir*		39.1529	-120.9374	
5	New Melones*		37.9917	-120.5337	
5	Paradise Lake*		39.8555	-121.5748	

 Table 1. Study Lakes by Water Quality Control Board (RWQCB).

		Chlorophyl-a			Epilimnion	Hypolimnion	THg in water	THg in sediment	
RWQCB	Lake	Station	(mg/L)	DOC (mg/L)	SO4 (mg/L)	MeHg (ng/L)	MeHg (ng/L)	(ng/L)	(Dry, ug/g)
1	Sonoma	Smith Creek	1.36	9.03	7.39	0.033	0.040	0.920	0.171
1	Sonoma	Dam	0.82	9.81	7.39	0.032	0.035	1.666	0.254
1	Mendocino	Dam	1.79	10.14	8.13	0.052	0.049	1.213	0.100
1	Mendocino	Russian River	2.02	8.88	8.14	0.047	0.043	1.231	0.045
1	Pillsbury		1.92	11.60	5.85	0.158	0.135	1.032	0.195
3	Naciemento	Dam	1.66	8.27	38.29	0.043	0.116	1.012	0.048
3	Naciemento	Las Tablas	5.41	12.85	40.57	0.215	NA	14.708	3.090
3	San Antonio	Marina	7.02	18.97	67.43	0.045	0.351	0.971	0.067
3	San Antonio	Delta	13.86	17.15	68.29	0.046	NA	1.342	0.076
5	Lake Engelbright		0.44	1.92	4.45	0.042	0.045	0.850	0.214
5	Thermalito Afterbay		1.38	2.03	3.90	0.025	NA	0.683	0.012
5	Thermalito Forebay		1.80	1.93	3.75	0.022	0.010	0.792	0.051
5	Lake Oroville		1.58	1.91	3.85	<rl< th=""><th>0.026</th><th>0.506</th><th>0.066</th></rl<>	0.026	0.506	0.066
5	Folsom Lake		1.17	2.02	3.00	0.043	0.034	0.644	0.126
5	Lake Natomas		1.18	1.94	3.03	0.033	0.035	0.919	0.055
5	Don Pedro		0.91	1.47	2.10	0.032	0.026	0.266	0.128
5	Lake McClure		1.33	1.77	3.00	0.041	0.025	0.354	0.081
5	Lake McSwain		1.63	2.08	2.72	0.033	0.029	1.552	0.115
5	San Luis Resrvoir		6.01	11.66	43.17	0.043	0.032	0.442	0.071
5	Oneil Forebay		1.98	8.62	47.80	0.043	0.042	0.803	0.105
5	Camp Far West		2.04	3.97	7.85	0.067	0.069	2.580	0.599
5	Bass Lake*		NC	NC	NC	<rl< th=""><th><rl< th=""><th>NC</th><th><rl< th=""></rl<></th></rl<></th></rl<>	<rl< th=""><th>NC</th><th><rl< th=""></rl<></th></rl<>	NC	<rl< th=""></rl<>
5	Lake Britton*		NC	NC	NC	<rl< th=""><th><rl< th=""><th>NC</th><th>0.030</th></rl<></th></rl<>	<rl< th=""><th>NC</th><th>0.030</th></rl<>	NC	0.030
5	Butt Valley Reservoir*		NC	NC	NC	<rl< th=""><th><rl< th=""><th>NC</th><th>0.120</th></rl<></th></rl<>	<rl< th=""><th>NC</th><th>0.120</th></rl<>	NC	0.120
5	Rollins Reservoir*		NC	NC	NC	0.042	0.037	NC	0.804
5	New Melones*		NC	NC	NC	<rl< th=""><th><rl< th=""><th>NC</th><th>0.098</th></rl<></th></rl<>	<rl< th=""><th>NC</th><th>0.098</th></rl<>	NC	0.098
5	Paradise Lake*		NC	NC	NC	<rl< th=""><th><rl< th=""><th>NC</th><th>0.147</th></rl<></th></rl<>	<rl< th=""><th>NC</th><th>0.147</th></rl<>	NC	0.147
8	Big Bear		3.80	13.73	20.83	0.032	<rl< th=""><th>0.432</th><th>0.066</th></rl<>	0.432	0.066
8	Irvine	Dam	5.79	44.66	235.00	0.090	0.322	0.767	0.192
8	Irvine	Santiago Flats	5.87	45.14	229.00	0.141	0.166	1.671	0.186
8	Perris	Dam	5.06	14.22	51.67	0.039	0.031	0.234	0.061
8	Perris	Allesandro	5.58	14.25	52.67	0.036	NC	0.272	0.045
8	Hemet		4.87	10.89	15.80	0.030	0.196	0.278	0.037
8	Elsinore		42.64	65.35	220.00	0.042	0.076	1.098	0.029

Table 2. Average of anlytes collected at each study station. *Sampled only once. NC = No Collection and <RL = below reporting limit.

			Normalized Tissue Hg	Average Aqueous	Geomtric Mean Aqueous		Geometric
RWQCB	Lake	Spp. Collected	(ng•g ⁻¹)	MeHg (ng•L ⁻¹)	MeHg (ng•L ⁻¹)	Average BAF	Mean BAF
1	Sonoma**	Largemouth Bass	677	0.030 ± 0.014	0.027	2.25×10^7	2.55×10^7
1	Mendocino**	Largemouth Bass	543	0.032 ± 0.025	0.023	1.71×10^7	2.33×10^7
1	Pillsbury**	Largemouth Bass	1314	0.158 ± 0.072	0.143	8.31×10^6	9.21×10^6
3	Naciemento**	Smallmouth Bass	1236	0.120 ± 0.099	0.079	1.03×10^7	1.57×10^7
3	San Antonio**	Largemouth Bass	302	0.146 ± 0.267	0.058	2.07×10^6	5.23×10^6
5	Camp Far West	Spotted Bass	843	0.068 ± 0.009	0.067	1.25×10^7	1.26×10^7
5	Lake Engelbright	Spotted Bass	521	0.036 ± 0.021	0.029	1.45×10^7	1.82×10^7
5	Thermalito Afterbay	Largemouth Bass	211	0.010 ± 0.009	0.010	1.22×10^7	1.35×10^7
5	Thermalito Forebay		NC	0.010 ± 0.005	0.010		
5	Lake Oroville	Smallmouth Bass	513	0.010 ± 0.005	0.010	4.53×10^7	4.74×10^7
5	Folsom Lake	Largemouth Bass	471	0.036 ± 0.024	0.031	1.3×10^7	1.53×10^7
5	Lake Natomas	Largemouth Bass	542	0.024 ± 0.013	0.020	2.27×10^7	2.7×10^7
5	Don Pedro Reservoir	Largemouth Bass	442	0.010 ± 0.008	0.010	3.36×10^7	3.7×10^7
5	Lake McClure	Largemouth Bass	769	0.023 ± 0.014	0.010	3.31×10^7	5.9×10^7
5	Lake McSwain	Largemouth Bass	535	0.025 ± 0.010	0.025	2.1×10^7	2.12×10^7
5	San Luis Resrvoir	Largemouth Bass	564	0.030 ± 0.016	0.026	1.87×10^7	2.2×10^7
5	Oneil Forebay	Largemouth Bass	234	0.040 ± 0.015	0.036	5.91 x 10 ⁶	6.54×10^6
5	Bass Lake*	Largemouth Bass	91	0.021	0.010	4.46×10^6	5.75×10^6
5	Lake Britton*	Smallmouth Bass	248	0.010	0.010	2.48×10^7	2.48×10^7
5	Butt Valley Reservoir	* Smallmouth Bass	180	0.010	0.010	1.8×10^7	1.8×10^7
5	Rollins Reservoir*	Smallmouth Bass	762	0.039	0.038	1.96 x 10 ⁷	2.01×10^7
5	New Melones*	Largemouth Bass	1125	0.010	0.010	7.6×10^7	8.81×10^7
5	Paradise Lake*	Largemouth Bass	161	0.010	0.010	1.61×10^7	1.61×10^7
8	Big Bear	Largemouth Bass	178	0.010 ± 0.012	0.010	9.2 x 10 ⁶	1.09×10^7
8	Irvine**	Largemouth Bass	479	0.161 ± 0.162	0.115	2.98×10^6	4.18×10^6
8	Perris**	Largemouth Bass	98	0.023 ± 0.015	0.010	4.25×10^6	5.19 x 10 ⁶
8	Hemet	Largemouth Bass	166	0.087 ± 0.202	0.032	1.91 x 10 ⁶	5.18×10^6
8	Elsinore	Largemouth Bass	121	0.054 ± 0.035	0.047	2.23 x 10 ⁶	2.56 x 10 ⁶

Table 3. *Lakes were sampled once in September 2009 as part of RWQCB 5's TMDL program. ** Multiple stations within lake sampled for water and sediment. (NC= No calculation) No fish were collected at Thermalito Forebay. Values below the reporting limit (RL) were set to 1/2 the RL (0.010 ng/L). Data presented here are lake-wide averages of methylmercury in water (Value \pm S.D.). Tissue tissue mercury values are normalized (350 mm) and represent the lake-wide average.



Figure 1. Median, upper and lower quartiles (25th and 75th percentile), standard deviation, and outliers (90th percentile) for the 21 study lakes.Observed aqueous methylmercury (MeHg) concentrations in stratified and non-stratified lake regimes (a) and the distribution of MeHg in stratified (summer) lakes (b). Stratified regimes were more variable due to the larger number of outliers (spikes in MeHg) especially in bottom water MeHg concentrations.



Figure 2. MeHg profiles of Lake San Antonio(a) and Lake Nacimiento(b) collected in September 2009. Lake San Antonio had relatively low aqueous MeHg concentrations in near surface and elevated values in bottom water that was a typical and consistent observation throughout the study period. Lake Nacimiento was relatively consistent in both near surface and bottom water and consistent with the observed data from the study (Not illustrated here). However, there was a large concentration of aqueous MeHg was around 12 m and was considered to be associated with either the thermocline or oxycline or both.

		X C	ŝ.	or .	5H	sp ^C			x	N ⁱ N	18 m	n ^a st	18 bm	واه
	LMB	ANG.	ANG.	ÁNG.	* NVG.	Chl-a	poc	50 ⁴	THS	TMM	THS	TMM	TH8-	NOC'
Tissue	<u> </u>	<u> </u>	<u> </u>	V.	V.		v	Ý	7	,	,	,	7	
AVG T	0.18528	1												
AVG_O2	0.06041	-0.39525	1											
AVG_pH	-0.36265	0.09405	-0.25662	1										
AVG_spC	-0.61586	0.23645	-0.37226	0.78898	1									
Chl-a	-0.61251	0.27231	-0.4296	0.47082	0.81025	1								
DOC	-0.53898	0.34632	-0.45328	0.79274	0.97301	0.83751	1							
SO4	-0.69512	0.15617	-0.38392	0.67829	0.96627	0.89434	0.94588	1						
THg_tw	0.36848	0.09893	-0.49394	0.23302	0.11339	0.08815	0.23553	0.0962	1					
TMMHg_tw	0.47314	0.39861	-0.31803	0.25632	0.12485	-0.02341	0.18591	0.02414	0.58108	1				
THg_Bw	0.36526	-0.33559	-0.2199	-0.04951	-0.122	-0.09084	-0.10915	-0.11006	0.62041	0.38467	1			
TMMHg_bw	-0.02954	0.30264	-0.32969	0.28866	0.33369	0.38897	0.4299	0.37585	0.33498	0.52807	0.22737	1		
THg_s	0.70944	-0.14146	0.0953	-0.30631	-0.46092	-0.54687	-0.415	-0.49307	0.48343	0.42004	0.61269	0.01337	1	
TOC%	-0.43652	-0.32276	0.15484	0.02693	0.29186	0.36856	0.18224	0.35519	-0.24156	-0.36333	0.1654	-0.05446	-0.07641	1
(b)														
		.5	¢	J	A	C					. 0			
	NB	113 x1Q) NG.	yr 10,	NG.	58 222-0	o ^C	0 ^k	128	MM	15 18		0	
	T _M ,	Þ.	A.	A	A	CN	$\mathcal{V}^{\mathbf{c}}$	ŞU	- V	1st	- TV-	Ń		
Tissue	1													
AVG_T	0.19466	1												
AVG_O2	0.01918	-0.53182	1											
AVG_pH	-0.36265	0.05279	-0.25427	1	_									
AVG_spC	-0.62083	0.16736	-0.22832	0.7961	1									
Chl-a	-0.60778	0.24611	-0.21666	0.66613	0.85019	1								
DOC	-0.31898	0.18126	-0.63398	0.78951	0.74344	0.67287	1							
SO4	-0.56732	0.23578	-0.2041	0.66279	0.95235	0.8269	0.67147	1						
THg	0.43282	-0.1706	0.46104	-0.13293	-0.22685	-0.12763	-0.12397	-0.17853	1					
ТММНд	0.23537	0.27531	-0.16781	0.563	0.39568	0.44563	0.53257	0.37381	0.30731	1				
THg_s	0.70944	-0.13096	0.21929	-0.30631	-0.45621	-0.52197	-0.23493	-0.36488	0.51263	0.31606	1			
TOC%	-0.43652	-0.33809	-0.09741	0.02693	0.26883	0.23883	0.23352	0.18961	-0.30104	-0.07813	-0.07641	1		

Table 4. (a)Correlation matrix of summer variables (r). (b)Winter variables. Values highlighetd in bold are significant (p<0.05; LMB only)



Figure 3. Logarithmic function of total mercury in largemouth bass expressed as a logarithmic function of total mercury in sediments.



Table 2. The table is sorted ascending on RWQCB. Darker shades represent higher average concentrations during the summer period relative to the rest of the study lakes. $\Delta O2$ represents a percent change over time of the oxycline. In general, the longer a lake maintains an oxycline, coupled with the difference between O2 max and min, the greater the percent change. General areas of lakes: RWQCB 1 = Coast Range, RWQCB 5 = Sierra Nevada, RWQCB 8 = Southern California



Figure 4. Delta O2 over time versus mercury in near surface water for RWQB 5 lakes (Sierra Nevada Region).